

# Recurrent Encephalopathy: NAGS (N-Acetylglutamate Synthase) Deficiency in Adults

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**ABSTRACT:** N-acetyl-glutamate synthase (NAGS) deficiency is a rare autosomal recessive urea cycle disorder (UCD) that uncommonly presents in adulthood. Adult presentations of UCDs include; confusional episodes, neuropsychiatric symptoms and encephalopathy. To date, there have been no detailed neurological descriptions of an adult onset presentation of NAGS deficiency. In this review we examine the clinical presentation and management of UCDs with an emphasis on NAGS deficiency. An illustrative case is provided. Plasma ammonia levels should be measured in all adult patients with unexplained encephalopathy, as treatment can be potentially life-saving. Availability of N-carbamylglutamate (NCG; carglumic acid) has made protein restriction largely unnecessary in treatment regimens currently employed. Genetic counselling remains an essential component of management of NAGS.

**RÉSUMÉ:** Encéphalopathie récurrente : le déficit en NAGS (N-acétylglutamate synthase) chez l'adulte. Le déficit en N-acétyl-glutamate synthase (NAGS) est un désordre rare du cycle de l'urée (DCU) transmis de façon autosomique récessive qui survient rarement à l'âge adulte. Le mode de présentation des DCU chez l'adulte inclut des épisodes de confusion, des symptômes neuropsychiatriques et une encéphalopathie. À ce jour, il n'existe pas de description neurologique détaillée du mode de présentation du déficit en NAGS chez l'adulte. Dans cette revue, nous examinons le mode de présentation clinique et la prise en charge des DCU, particulièrement le déficit en NAGS, ainsi qu'une observation clinique. Les niveaux plasmatiques d'ammoniac devraient être mesurés chez tous les patients adultes présentant une encéphalopathie inexplicquée, étant donné que le traitement de cette pathologie peut leur sauver la vie. Depuis que le N-carbamylglutamate (NCG; acide carglumique) est disponible, il n'est plus nécessaire de restreindre l'apport en protéines chez ces patients. Le conseil génétique demeure une composante essentielle de la prise en charge du déficit en NAGS.

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In humans and other mammals nitrogen is produced by the catabolism of proteins and excreted as urea via the kidneys through the process of the urea cycle<sup>1</sup> (Figure 1). The urea cycle which is active primarily in the liver converts waste nitrogen into ammonium. Deficiencies of enzymes in the urea cycle may result in the accumulation of ammonium. Urea cycle disorders may present from infancy to adulthood.

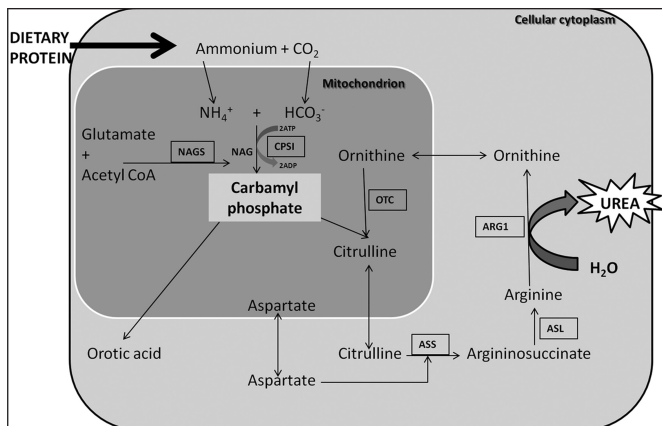
## Urea cycle review

The urea cycle consists of six enzymes and two mitochondrial membrane transporters (Figure 1): N-acetyl glutamate synthase (NAGS), carbamyl phosphate synthetase I (CPSI), ornithine transcarbamylase (OTC), argininosuccinate synthetase (ASS), argininosuccinate lyase (ASL), arginase, the aspartate transporter (citrin), and the ornithine transporter<sup>1-3</sup>. An interruption in any step of the urea cycle may cause a build-up of ammonium which is neurotoxic and if untreated may result in coma and death. All disorders except for X-linked OTC

deficiency are inherited in an autosomal recessive manner. Enzymes located in the mitochondria are CPSI, NAGS, and OTC; these can therefore be affected by other mitochondrial diseases or perturbations. The incidence of urea cycle disorder (UCDs) in the United States is approximately 1 in 8200<sup>4</sup>. Prenatal testing based on mutation analysis is available for all six conditions<sup>5-7</sup>.

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**Figure 1:** Simplified version of urea cycle depicted. The urea cycle converts protein into urea which is excreted by the kidneys. There are six enzymes involved: N-acetylglutamate synthase (NAGS), carbamyl-phosphate-synthetase-I (CPSI), ornithine transcarbamylase (OTC), argininosuccinate synthetase (ASS), argininosuccinate lyase (ASL), arginase (ARG1). Further abbreviations: adenosine triphosphate (ATP), adenosine diphosphate (ADP). Redrawn with permission from: Lee B. Urea cycle disorders: Management. In: UpToDate, Basow, DS (Ed), UpToDate, Waltham, MA 2012. Copyright © 2012 UpToDate, Inc. For more information visit [www.uptodate.com](http://www.uptodate.com).

**Clinical presentation of UCDs**

The typical presentation for a urea cycle disorder occurs in the first few days of life. The infant may present with gastrointestinal symptoms such as vomiting occurring after feeding (protein load). Neurological symptoms such as lethargy, seizures and coma can follow quickly, a presentation identical to that of an infant with sepsis. A non-diagnostic work-up for sepsis should raise the clinical suspicion for an inborn error of metabolism. A common sign is hyperventilation and respiratory alkalosis as ammonium is a central nervous system (CNS) stimulant. Hyperventilation is thought to result from cerebral edema caused by the build up of ammonium;<sup>8</sup> however hyperventilation can also be seen without evidence of cerebral edema. Neonates who present in the first few days of life do so as a result of the catabolic stress of labour and delivery and low fluid intake in the immediate post natal period.

Patients who have partial enzyme deficiencies, such as female carriers (X-linked OTC deficiency) will often have a delayed presentation despite a lifelong history of chronic cyclical nausea and vomiting, and possibly a seizure disorder or a psychiatric illness<sup>9</sup>. There may also be developmental delay. Many patients self-select a low protein diet. In all groups of patients hyperammonemic crises may occur with increased catabolic stress caused by infection, starvation, surgery or trauma.

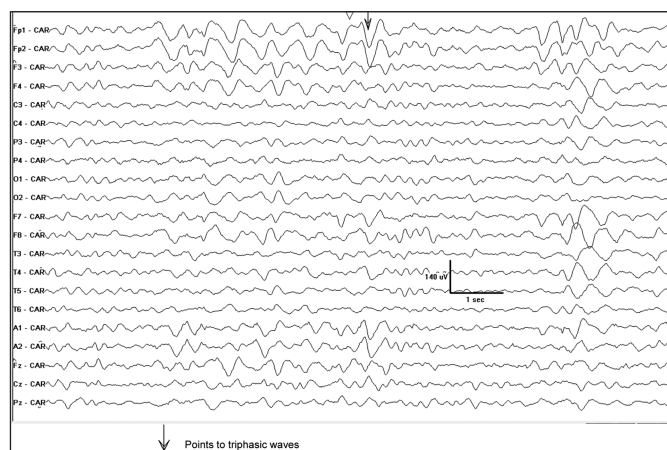
**Approach to a patient with a suspected UCD**

The approach to a patient considered to have a UCD includes a comprehensive neurological assessment with particular attention to family history and key clinical features such as behavioural changes, protein aversion and gastrointestinal symptoms. Investigations should first and foremost include an ammonia level. Other key investigations include arterial pH,

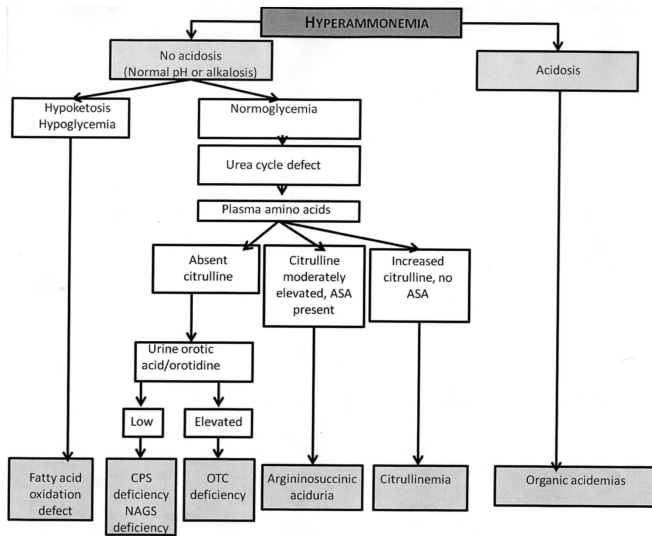
- Complete blood count with differential
- Urinalysis
- Blood gases
- Serum electrolytes
- Liver enzymes and liver function tests
- Blood glucose
- **Plasma ammonia**
- Urine ketones if acidosis or hypoglycemia present
- Quantitative plasma and urine amino acids
- Urine organic acids
- Urine orotic acid
- Plasma lactate
- Carnitine (acyl and free)
- Acylcarnitine profile
- DNA for banking
- Liver biopsy for enzyme studies (now only rarely required due to easy availability of DNA based mutation studies)

**Figure 2:** Recommended laboratory studies for a patient with a suspected inborn error of metabolism. Modified from Burton B. Inborn errors of Metabolism in Infancy: A Guide to Diagnosis. Pediatrics 1998;102(6):e69-e77.

serum lactate, serum glucose and cerebrospinal fluid (CSF) analysis, (with hyperammonemia there can be cerebral edema so CSF analysis should only be performed with caution) (Figure 2). It is important to note that hyperammonemia may be chronic or occur only during metabolic decompensation and therefore investigations can be normal<sup>10</sup>. Electroencephalogram (EEG) (Figure 3) and magnetic resonance (MR) imaging can be helpful investigations. Fluid attenuated inversion recovery sequence (FLAIR) on MR imaging has been suggested to identify white matter tract abnormalities that can exist in UCDs<sup>11</sup>. Other imaging techniques described to show abnormalities include diffusion tensor imaging (DTI) and MR spectroscopy. If plasma ammonia is elevated then metabolic indices such as plasma amino acids, urine orotic acid and urine organic acids should be measured. The importance of testing these metabolic indices is to differentiate among the various UCDs (Figure 4). A genetics and metabolic consultation is useful to proceed with further



**Figure 3:** Electroencephalogram. Referential montage (common average reference) shows diffuse background slowing (delta) with the intermittent appearance of triphasic waves, with a bifrontal predominance.



**Figure 4:** Flow chart for the approach to hyperammonemia. Modified from Burton B. *Inborn errors of Metabolism in Infancy: A Guide to Diagnosis*. Pediatrics 1998;102(6):e69-e77., and Summar M. *Current strategies for the management of neonatal urea cycle disorders*. J Pediatr. 2001 Jan;138(1 Suppl):S30-9.

work up for molecular analysis or tissue enzyme analysis. Patients suspected to be having a UCD should be managed in an intensive care setting as this is a medical emergency. Cerebral edema occurs early with severe hyperammonemia, and delays in reducing the level of ammonia may lead to serious neurological complications including death due to increased intracranial pressure with herniation. Survival from a severe episode often carries with it irreversible brain damage. Treatment should thus not be delayed if a UCD is suspected.

### Medical and metabolic management

In an emergent presentation, physiologic stabilization is most important; adjunctive treatment may include intravenous fluids, and possibly hemodialysis for ammonia removal. The mainstay of ongoing management of NAGS deficiency, as with all UCDs is maintenance of plasma ammonia levels in a normal range by limiting protein intake, avoiding periods of catabolic stress, and using nitrogen scavenger drugs to allow an alternate pathway for the excretion of nitrogen precursors<sup>12,13</sup> (e.g. sodium phenylbutyrate). Recently, in NAGS deficiency, specific treatment with carglumatic acid, a structural analog of NAG has been used as it has been shown to activate CPSI<sup>14-17</sup> and restore ureagenesis<sup>2,9</sup>. Patients receiving carglumatic acid typically do not require a protein restricted diet.

### NAGS deficiency

N-acetylglutamate synthase (NAGS; MIM# 608300), one of the three mitochondrial enzymes of the urea cycle, produces N-acetylglutamate (NAG) from glutamate and acetyl coenzyme A (Acetyl CoA). N-acetylglutamate was first identified in the 1950's and initially discovered as an intermediate in the arginine-biosynthetic pathway of *Escherichia coli*. N-acetyl glutamate synthase was later described as an essential allosteric cofactor of

mitochondrial carbamylphosphate synthetase I (CPSI), the first enzyme of the urea cycle<sup>18</sup>. N-acetyl glutamate synthase is primarily expressed in the liver and in the small intestine and its product, NAG, is postulated to activate CPSI<sup>19</sup>. Hyperammonemia can result once CPSI is deprived of its co-factor NAG.

The first case report of NAGS deficiency was published in 1981<sup>20</sup> and subsequently approximately 34 other cases of NAGS deficiency have been reported with most patients presenting in the neonatal period<sup>17</sup>. N-acetyl glutamate synthase deficiency is the least common UCD and few late-onset neurological presentations have been described<sup>2,20-22</sup>; the gene mutation maps to chromosome 17q21.31<sup>19</sup>. There are 22 published mutations up to date that have been described for NAGS<sup>3,23</sup>.

Deficiencies of NAGS activity can either be inherited (mutation in NAGS gene) or acquired by a secondary inhibition of NAGS activity in some conditions which cause short chain fatty acid accumulation such as some organic acidemias and the use of valproic acid.

A detailed summary of published cases of NAGS in the literature is listed in the Table. Literature searches were done using online data bases such as PUBMED, GENE REVIEWS and OMIM. In total, 34 cases of confirmed NAGS deficiency were identified. This mini review will help with the understanding of the genetic aspects and key neurologic features associated with UCDs and therefore prompt earlier diagnosis and treatment. We discuss the unique presentation of an adult-onset UCD. Our patient had a history of behavioural change and confusion which is in keeping with a late-onset UCD. An adult presentation of NAGS deficiency however is very rare. No trigger was identified in our patient to account for his hyperammonemic crises.

### Illustrative Case

The patient was symptomatic in early childhood and adulthood with symptoms of hyperammonemia in the setting of a negative family history although no ammonia level was checked until his hospitalization at age 38 years. A key clinical feature was an almost 20 year history of fluctuating behavioural changes associated with nausea and vomiting. A 38-year-old left-handed dairy farmer was referred for acute confusion and bizarre behavioural changes. His past medical history was notable for similar episodes dating as far back as 18 years. There had been no previous psychiatric or neurologic evaluation, however, a normal cranial computed tomogram (CT) scan had been reported. He was taken to the emergency room by his wife and upon assessment he described ongoing nausea and vomiting as well as headache. The patient had completed secondary school and two years of college with no history of learning difficulties. He was from a non-consanguineous Irish and Scottish background with no relevant family history.

On examination vital signs were stable. There was behavioural disinhibition and fluctuating drowsiness. A minimal status examination (MMSE) score was 23/30, (points lost for five-minute recall, three-step command, and constructing intersecting pentagons). Cranial nerve examination and power testing was normal. There was mild spasticity on assessment of muscle tone with brisk reflexes, sustained ankle clonus, and down-going plantar reflexes. Coordination was impaired.

**Table: Summary of findings in reported cases of confirmed N-acetylglutamate synthase (NAGS) deficiency**

Case	Age at diagnosis; Sex	Background Consanguinity	Family History	Clinical findings	Death	Peak Ammonia* mmol/L	Clin on admission* Normal 109-150 mmol/L	Cit on admission* Normal 10-30 mmol/L	Diagnosis by: enzyme analysis OR molecular studies	Citation
1	6 male	Ukn	Ukn	vomiting, feeding intolerance, episodic confusion	No	500	Ukn	Ukn	NAGS deficiency on mutation analysis	Come et al. Mol Genet Metab 2011; 102:275. <sup>31</sup>
2	Screened at birth; female	Turkish	Yes; older sister died of severe hyperammonemia (NAGS deficiency not confirmed)	asymptomatic	No	393	Ukn, but 1961 on subsequent admission	Ukn	Second liver biopsy: reduced NAGS activity, later found to be homozygous for NAGS mutation	Gesser et al. Eur J Pediatr 2010; 169:197-199. <sup>32</sup>
3	41 birth; Ukn	First cousins	Ukn	coma, seizures	No	>1400	Ukn	Ukn	NAGS deficiency on mutation analysis	Nordenstrom et al. J Inher Metab Dis 2007; 30:400. <sup>36</sup>
4	40 years; female	Ukn	Ukn	intermittent staring spells, nausea, recurrent vomiting, lethargy, ataxia, migraine headaches, eventually coma	No	500	Ukn	Ukn	NAGS deficiency on mutation analysis	Tuchman et al. Pediatr Res 2008; 64(2):213-217. <sup>22</sup>
5	33 years; female	Ukn	Ukn	episodic altered mental status with coma after cesarean section. Psychomotor seizures	No	138 - 4781	Ukn	Ukn	NAGS deficiency on mutation analysis	Grody et al. J Inher Metab Dis 1994; 17(5):566-574. <sup>37</sup>
6	4 days; female	Ukn	Ukn	hypotonia, drowsiness, tremor, and no suck	No	182	1150	13	NAGS deficiency on mutation analysis	Caldovic et al. Hum Mutat. 2007; 28(8):754-759. <sup>3</sup>
7	3 months; male	French	Ukn	muscular axial hypotonia, hyporeactivity, hepatomegaly	No	113-367	841	19	NAGS deficiency on mutation analysis	Guffon et al. J Pediatr 2005; 147:260-262. <sup>38</sup>
8	Screened at birth; male	Ukn	Older sister died of severe hyperammonemia, no diagnosis made	asymptomatic	No	208	1114	<4	NAGS deficiency on mutation analysis	Schmidt et al. J Pediatr 2005; 147:260-262. <sup>38</sup>
9	4 weeks; female	Ukn	No	recurrent vomiting, irritability, lethargy, later-on headaches, hallucinations	No	256	elevated	normal	Initially NAGS not assayed on liver biopsy	Caldovic et al. J Pediatr 2004; 145:552-554. <sup>41</sup>
10	9 years; female	White	Yes; younger sister case 9	lethargy, anorexia, vomiting, respiratory distress and seizures	No	978	1710	ND	Later NAGS deficiency on mutation analysis	Caldovic et al. Hum Mutat. 2003; 25(3):298. <sup>2</sup>
11	35 years; male	No	None	post-operative combativeness, confusion, seizures	Yes	621	elevated	Low	NAGS deficiency on mutation analysis	Belanger-Quintana et al. Eur J Pediatr. 2003; 162(11):773-775. <sup>40</sup>
12	12 years; male	Ukn	Ukn	lethargy, decreased LOC with protein, EEG showed left temporal lobe spikes	No	350	801	21	Liver biopsy: L-Arginine unable to activate NAGS and normal CPS activity.	Haberle et al. Human Mutat. 2003; 21(6):593-597. <sup>23</sup>
13	3 days; Ukn	Turkish	Ukn	Ukn	Ukn	Ukn	Ukn	Ukn	NAGS deficiency on mutation analysis	Heckmann et al. Acta Paediatr. 2005; 94(1):121-124. <sup>41</sup>
14	3-4 days; Ukn	Turkish	Ukn	Ukn	Ukn	Ukn	Ukn	Ukn	NAGS deficiency on mutation analysis	Haberle et al. Human Mutat. 2003; 21(6):593-597. <sup>23</sup>
15	6 days; Ukn	German	Ukn	Ukn	Ukn	Ukn	Ukn	Ukn	Liver biopsy: No NAGS activity; later NAGS mutation found	Haberle et al. Human Mutat. 2003; 21(6):593-597. <sup>23</sup>
16	3 days; Ukn	Turkish	Ukn	Ukn	Ukn	Ukn	Ukn	Ukn	NAGS deficiency on mutation analysis	Haberle et al. Human Mutat. 2003; 21(6):593-597. <sup>23</sup>
17	3 days; female	German	Ukn	signs of cerebral edema	Yes	942			Liver biopsy: reduced NAGS activity, NAGS mutation confirmed using cultured fibroblasts and DNA sequencing	Schmidt et al. Biochim Biophys Acta. 2005; 1740:54-59. <sup>39</sup>
18	16 hours; male	Faeroe Islands	Ukn	lethargy, jitteriness, hypertonia, hypothermia, coma	Yes	Ukn	elevated	ND	Post-mortem liver tissue: reduced NAGS	Caldovic et al. Hum Genet 2003; 112:364-368. <sup>42</sup>
19	3 days; female	Distant approximately 5 generations back	No	lethargy, anorexia, respiratory distress, coma, seizures	No	1700	1268	ND	Later parents found to be heterozygotes	Takamashi J., et al. Am J Neuroradiol (in press) 2002. <sup>44</sup>
20	2 days; female	Hipaun/No	Yes; younger sibling case 19	lethargy, anorexia, vomiting, respiratory distress and seizures	No	978	1710	ND	Molecular studies: both homozygous for NAGS gene mutation	Caldovic et al. Hum Genet 2003; 112:364-368. <sup>42</sup>
21	4 days; male	Iranian Jewish/First cousins	Ukn	unresponsive, seizures	No	1300	2652	ND	Initial liver biopsy inconclusive, later molecular analysis confirmed homozygous NAGS gene mutation	Elpeleg et al. Am Neurol. 2002; 52(6):845-89. <sup>45</sup>
22	at birth; male	Ukn	Yes; younger brother to case 21	coma	No	1900	1732	ND	Homozygous NAGS gene mutation	Fogel et al. Acta Paediatr. 1999; 88(12):1409-1411. <sup>46</sup>
23	4 years; male	Ukn	None	irritability, vomiting, lethargy, protein refusal, ataxia, worsened with VPA	No	229	920	Ukn	Liver biopsy: NAGS activity reduced 26% control	Piecko et al. Eur J Pediatr. 1998; 157(12):996-998. <sup>47</sup>
24	12 years 11 months; female	Austrian/Slovenian	No	vomiting, protein aversion, restlessness, disorientation, aggression, hyperreflexia	No	221	1616	12	Partial NAGS deficiency Liver biopsy: NAGS activity reduced 15% control	Haberle et al. Hum Mutat. 2003; 21(6):593-597. <sup>23</sup>

25	4 days; female	Ukn	Ukn	Ukn	316	1186	ND	ND	Liver biopsy: NAGS activity <5% control	Morris et al. J. Inher. Metab. Dis. 1998;21(8):867-868. <sup>48</sup>
26	20 years; male	Ukn	Ukn	Ukn	525	1571	2	No	Partial NAGS deficiency Liver biopsy: NAGS activity <50% control	Himie et al. J. Inher. Metab. Dis. 1997;20(6):839-840. <sup>49</sup>
27	3-4 days; male	Ukn	Ukn	Ukn	530	1054	<1	No	Liver biopsy: NAGS activity <10% control	Guffon et al. J. Inher. Metab. Dis. 1995;18(1):61-65. <sup>17</sup>
28	5 years; 7 months; male	Ukn	Ukn	Ukn	>200	850	24	No	Liver biopsy: NAGS activity reduced to 9.7% of control	Vockley et al. Biochem. Med. Metab. Biol. 1992;47(1):38-46. <sup>5</sup>
29	5 months; female	Ukn	Ukn	Ukn	285	755	6	No	Partial NAGS deficiency Liver biopsy: NAGS activity reduced to 40% of control	Burlin et al. J. Inher. Metab. Dis. 1992;15(3):395-398. <sup>51</sup>
30	5.6 weeks; male	Ukn	Ukn	Ukn	215	Elevated	Low	No	Liver biopsy revealed NAGS activity undetectable	Pondy et al. J. Inher. Metab. Dis. 1991;14(5):685-690. <sup>52</sup>
31	26 days; male	Ukn	Ukn	Ukn	185	Elevated	Low	No	None: positive family history	
32	13 months; female	Muslim	Ukn	Ukn	241	549	ND	Yes	Partial NAGS deficiency Liver biopsy: NAGS activity reduced to 33% of control	Elpeck et al. Eur. J. Pediatr. 1990 Jun;149(9):654-656. <sup>53</sup>
33	6 days; male	Ukn	Ukn	Ukn	711	940	ND	Yes	Liver biopsy: NAGS activity Not detectable	Bachmann et al. J. Inher. Metab. Dis. 1988;11(2):191-193. <sup>54</sup>
34	Screened at 3 days; male	Ukn	Ukn	Ukn	242	1020	Ukn	Died at age 9	Liver biopsy: NAGS activity Not detectable	Bachmann et al. N. Engl. J. Med. 1981 Feb 26;304(9):544. <sup>50</sup> (INDEX CASE) Schubiger et al. Eur. J. Pediatr. 1991 Mar;150(5):353-356. <sup>55</sup>
35	38; male	Irish-Scottish	Ukn	Ukn	434	1062	15	No	NAGS deficiency on mutation analysis	

Ukn (unknown), LOC (loss of consciousness), CPS (carbamoyl phosphate synthetase) mat. (maternal), EEG (electroencephalogram), VPA (valproic acid), Gln (Glutamine), Cit (Citrulline), ND (Not Detectable)

Significant asterixis was present and gait assessment was normal. Otherwise general examination was unremarkable.

The patient was admitted to hospital. Extensive laboratory studies were all within normal limits. Magnetic resonance imaging of head and full-spine were reviewed with expert neuro-radiologists and deemed normal. Plasma ammonia was found to be markedly elevated at 434 umol/L (15-55 umol/L). A blood gas revealed a mild respiratory alkalosis. Continuous EEG monitoring (Figure 3) demonstrated severe generalized encephalopathy with associated triphasic waves suggesting metabolic or hepatic etiology with no epileptiform activity.

The patient responded well to intravenous fluids, lactulose and a relatively lower protein diet. Upon discharge plasma ammonia levels decreased to 85 umol/L. A referral was made to a genetics/metabolics specialist to further investigate an underlying metabolic etiology.

A complete metabolic work-up revealed an elevated glutamine level of 1062 µmol/L (109-750 µmol/L), normal citrulline at 15 µmol/L (10-50 µmol/L), and normal urine amino acids. Urine orotic acid level was normal. On molecular studies OTC gene sequencing was normal (this was done as OTC deficiency is the most common UCD and orotic acid can be normal in OTC patients). Molecular sequencing of the NAGS gene was performed and the patient was found to be a compound heterozygote for E433G and IVS6+5 G > A, both novel mutations. The intronic mutation involved a consensus base pair located 5 bp downstream of an exon/intron boundary, in the donor splice site of intron 6. A G>A substitution in this position is expected to reduce efficiency of splicing of NAGS mRNA and lead to decreased, but not absent, expression of NAGS from this allele<sup>24</sup>. The residual NAGS expression from this allele likely resulted in sufficient NAGS enzymatic activity to avoid neonatal hyperammonemia, and may serve to explain a delayed onset in adult life in our patient.

Mutation sequencing of NAGS was obtained in both parents. The patient's father was confirmed to be a carrier of the IVS6+5 G > A mutation, but interestingly, no mutation was identified in the patient's reported biological mother. Non-maternity, gonadal mosaicism or a *de novo* mutation may explain the absence of the E433G mutation in the mother.

**Medical management of our case**

A relatively low protein diet (around 1 gm/kg about <73 g/day) was commenced and our patient's ammonia level was maintained in the normal range. Initially he was also started on sodium phenylbutyrate 200 mg/kg TID (4500 mg TID) and citrulline 50mg/kg TID (1200 mg TID). Sodium phenylbutyrate reduces ammonia production by creating an alternate pathway for the excretion of nitrogen containing precursors while citrulline also aids in nitrogen clearance and in maintaining the arginine pool in proximal UCDs.

**Carglumic acid**

Our patient was eventually switched to carglumic acid 1200 mg TID and a more liberalized protein intake once the diagnosis of NAGS deficiency was confirmed. The switch was made to carglumic acid because of non-compliance with a low-protein diet, intolerance to sodium phenylbutyrate and citrulline (stomach distress and body odour) and the specificity of

carglumic acid to NAGS deficiency. Carglumic acid activates CPSI therefore leading to a reduction in ammonia levels. Currently our patient's ammonia levels are normal and range from 29-35  $\mu\text{mol/L}$ .

Carglumic acid is much more expensive than sodium phenylbutyrate (daily cost \$1961.00 Canadian dollars for carglumic acid vs. \$8.46 for sodium phenylbutyrate at current prices) however carglumic acid is the standard of care for patients with NAGS deficiency despite the cost, as treatment with scavengers and protein restriction are insufficient to prevent breakthrough hyperammonemia which would cause resultant increase in patient morbidity.<sup>25,26</sup>

### Long-term management and morbidity

Our patient did eventually resume his work on the farm but remains troubled with mild short term memory loss. His behaviour has also shown a marked improvement. He has not had a hyperammonemic crisis for the last two years. His therapeutic course has been plotted (Figure 5).

Patients with UCD can present at any age and during hyperammonemic crises mortality can be as high as 10%<sup>27</sup>. In chronic management, avoidance of periods of stress is important. Other neurologic sequelae of hyperammonemia may include seizures and/or developmental delay. Evidence suggests that virtually all survivors of a hyperammonemic coma are left with developmental delay<sup>28-30</sup>. Cognitive impairment in adult-onset presentations has been described even in asymptomatic OTC-deficient heterozygous women (learning disabilities and attention deficit hyperactivity disorder)<sup>31,32</sup>. Although not specific to NAGS deficiency a recent review by Gropman et al<sup>33</sup> does suggest that there is a neurochemical basis for cognitive and motor delay that may not only involve ammonia and glutamine as neurotoxins, but also alterations in the levels of

neurotransmitters all leading to neuropathological changes. No specific literature exists regarding documenting cognitive deficits in adult-onset presentation of NAGS deficiency as it is a very rare condition.

### CONCLUSIONS

Plasma ammonia should be measured in all patients with an unexplained encephalopathy including cyclical presentations to identify potential underlying metabolic disorders. Electroencephalogram can be a clue with the presence of triphasic waves indicating a metabolic encephalopathy. Hyperammonemia with a normal anion gap and normal serum glucose should be investigated to rule out a urea cycle disorder. Our illustrative case describes a rare presentation of a rare urea cycle disorder. Management for these patients requires a multidisciplinary approach including a dietician, social worker and a genetic counsellor. Awareness of inborn errors of metabolism presenting with neuro-psychiatric manifestations is essential for all adult neurologists and psychiatrists. Genetic counselling is an important component of UCDs management.

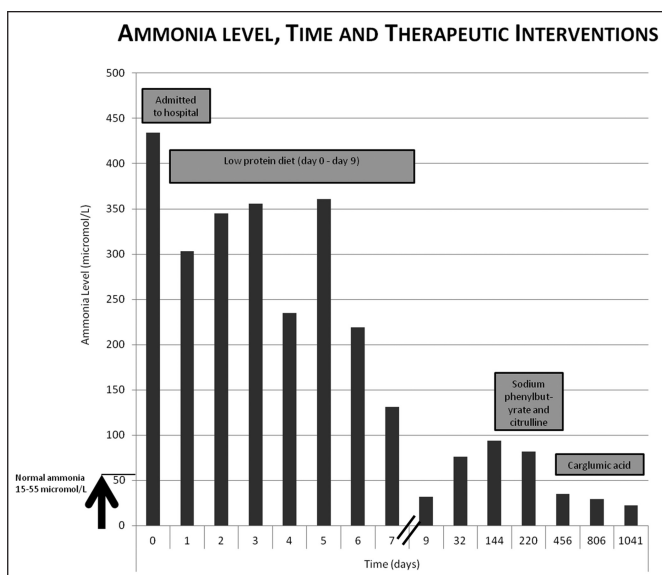
### ACKNOWLEDGEMENTS

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The NAGS mutation testing was done in Dr. Mendel Tuchman's laboratory in Washington, DC.

### REFERENCES

1. Brusilow SW. Urea cycle disorders: clinical paradigm of hyperammonemic encephalopathy. *Prog Liver Dis*. 1995;13:293-309.
2. Caldovic L, Morizono H, Panglao MG, et al. Late onset N-acetylglutamate synthase deficiency caused by hypomorphic alleles. *Hum Mutat*. 2005;25:293-8.
3. Caldovic L, Morizono H, Tuchman M. Mutations and polymorphisms in the human N-acetylglutamate synthase (NAGS) gene. *Hum Mutat*. 2007;28:754-9.
4. Brusilow SW, Maestri NE. Urea cycle disorders: diagnosis, pathophysiology, and therapy. *Adv Pediatr*. 1996;43:127-70.
5. Altarescu G, Brooks B, Eldar-Geva T, et al. Polar body-based preimplantation genetic diagnosis for N-acetylglutamate synthase deficiency. *Fetal Diagn Ther*. 2008;24:170-6.
6. Sniderman King L, Singh RH, Rhead WJ, Smith W, Lee B, Summar ML. Genetic counseling issues in urea cycle disorders. *Crit Care Clin*. 2005;21:S37-44.
7. Kamoun P, Fensom AH, Shin YS, et al. Prenatal diagnosis of the urea cycle diseases: a survey of the European cases. *Am J Med Genet*. 1995;55:247-50.
8. Butterworth RF. Effects of hyperammonemia on brain function. *J Inher Metab Dis*. 1998;21 Suppl 1:6-20.
9. Tuchman M, Lee B, Lichter-Konecki U, et al. Cross-sectional multicenter study of patients with urea cycle disorders in the United States. *Mol Genet Metab*. 2008;94:397-402.
10. Tuchman M, Yudkoff M. Blood levels of ammonia and nitrogen scavenging amino acids in patients with inherited hyperammonemia. *Mol Genet Metab*. 1999;66:10-15.
11. Gropman A. Brain imaging in urea cycle disorders. *Mol Genet Metab* 2010;100 Suppl 1:S20-30.
12. Batshaw ML, Painter MJ, Sproul GT, Schafer IA, Thomas GH, Brusilow S. Therapy of urea cycle enzymopathies: three case studies. *Johns Hopkins Med J*. 1981;148:34-40.
13. Batshaw ML, MacArthur RB, Tuchman M. Alternative pathway therapy for urea cycle disorders: twenty years later. *J Pediatr*. 2001;138:S46-54; discussion S54-5.



**Figure 5:** Graph of our patient's ammonia level versus time with therapeutic interventions included. Day 0 is day of presentation. Normal ammonia values 15-55  $\mu\text{mol/L}$ .

14. Ah Mew N, McCarter R, Daikhin Y, Nissim I, Yudkoff M, Tuchman M. N-carbamylglutamate augments ureagenesis and reduces ammonia and glutamine in propionic acidemia. *Pediatrics*. 2010; 126:e208-14.
15. Ah Mew N, Payan I, Daikhin Y, Nissim I, Tuchman M, Yudkoff M. Effects of a single dose of N-carbamylglutamate on the rate of ureagenesis. *Mol Genet Metab*. 2009;98:325-30.
16. Daniotti M, la Marca G, Fiorini P, Filippi L. New developments in the treatment of hyperammonemia: emerging use of carglumic acid. *Int J Gen Med*. 2011;4:21-8.
17. Guffon N, Vianey-Saban C, Bourgeois J, Rabier D, Colombo JP, Guibaud P. A new neonatal case of N-acetylglutamate synthase deficiency treated by carbamylglutamate. *J Inherit Metab Dis*. 1995;18:61-5.
18. Caldovic L, Morizono H, Daikhin Y, et al. Restoration of ureagenesis in N-acetylglutamate synthase deficiency by N-carbamylglutamate. *J Pediatr*. 2004;145:552-4.
19. Caldovic L, Ah Mew N, Shi D, Morizono H, Yudkoff M, Tuchman M. N-acetylglutamate synthase: structure, function and defects. *Mol Genet Metab*. 2010;100 Suppl 1:S13-19.
20. Bachmann C, Krahenbuhl S, Colombo JP, Schubiger G, Jaggi KH, Tonz O. N-acetylglutamate synthetase deficiency: a disorder of ammonia detoxication. *N Engl J Med*. 1981;304:543.
21. Bachmann C, Colombo JP, Jaggi K. N-acetylglutamate synthetase (NAGS) deficiency: diagnosis, clinical observations and treatment. *Adv Exp Med Biol*. 1982;153:39-45.
22. Tuchman M, Caldovic L, Daikhin Y, et al. N-carbamylglutamate markedly enhances ureagenesis in N-acetylglutamate deficiency and propionic acidemia as measured by isotopic incorporation and blood biomarkers. *Pediatr Res*. 2008;64:213-17.
23. Haberle J, Schmidt E, Pauli S, et al. Mutation analysis in patients with N-acetylglutamate synthase deficiency. *Hum Mutat*. 2003; 21:593-7.
24. Rogozin IB, Milanesi L. Analysis of donor splice sites in different eukaryotic organisms. *J Mol Evol*. 1997;45:50-9.
25. Ah Mew N, Caldovic L. N-acetylglutamate synthase deficiency: an insight into the genetics, epidemiology, pathophysiology, and treatment. *The Application of Clinical Genetics*. 2011;4:127-35.
26. Haberle J. Role of carglumic acid in the treatment of acute hyperammonemia due to N-acetylglutamate synthase deficiency. *Ther Clin Risk Manag*. 2011;7:327-32.
27. Batshaw ML, Msall M, Beaudet AL, Trojak J. Risk of serious illness in heterozygotes for ornithine transcarbamylase deficiency. *J Pediatr*. 1986;108:236-41.
28. Msall M, Monahan PS, Chapanis N, Batshaw ML. Cognitive development in children with inborn errors of urea synthesis. *Acta Paediatr Jpn*. 1988;30:435-41.
29. Uchino T, Endo F, Matsuda I. Neurodevelopmental outcome of long-term therapy of urea cycle disorders in Japan. *J Inherit Metab Dis*. 1998;21 Suppl 1:151-9.
30. Bachmann C. Long-term outcome of patients with urea cycle disorders and the question of neonatal screening. *Eur J Pediatr*. 2003;162 Suppl 1:S29-33.
31. Batshaw ML, Roan Y, Jung AL, Rosenberg LA, Brusilow SW. Cerebral dysfunction in asymptomatic carriers of ornithine transcarbamylase deficiency. *N Engl J Med*. 1980;302:482-5.
32. Gyato K, Wray J, Huang ZJ, Yudkoff M, Batshaw ML. Metabolic and neuropsychological phenotype in women heterozygous for ornithine transcarbamylase deficiency. *Ann Neurol*. 2004;55: 80-6.
33. Gropman AL, Batshaw ML. Cognitive outcome in urea cycle disorders. *Mol Genet Metab*. 2004;81 Suppl 1:S58-62.
34. Corne C, Fouilhoux A, Aquaviva C, Besson G. First French case of NAGS deficiency. 20 years of follow up. *Mol Genet Metab*. 2011;102:275.
35. Gessler P, Buchal P, Schwenk HU, Wermuth B. Favourable long-term outcome after immediate treatment of neonatal hyperammonemia due to N-acetylglutamate synthase deficiency. *Eur J Pediatr*. 2010;169:197-9.
36. Nordenstrom A, Halldin M, Hallberg B, Alm J. A trial with N-carbamylglutamate may not detect all patients with NAGS deficiency and neonatal onset. *J Inherit Metab Dis*. 2007;30:400.
37. Grody WW, Chang RJ, Panagiotis NM, Matz D, Cederbaum SD. Menstrual cycle and gonadal steroid effects on symptomatic hyperammonemia of urea-cycle-based and idiopathic aetiologies. *J Inherit Metab Dis*. 1994;17:566-74.
38. Guffon N, Schiff M, Cheillan D, Wermuth B, Haberle J, Vianey-Saban C. Neonatal hyperammonemia: the N-carbamoyl-L-glutamic acid test. *J Pediatr*. 2005;147:260-62.
39. Schmidt E, Nuoffer JM, Haberle J, et al. Identification of novel mutations of the human N-acetylglutamate synthase gene and their functional investigation by expression studies. *Biochim Biophys Acta*. 2005;1740:54-9.
40. Belanger-Quintana A, Martinez-Pardo M, Garcia MJ, et al. Hyperammonemia as a cause of psychosis in an adolescent. *Eur J Pediatr*. 2003;162:773-5.
41. Heckmann M, Wermuth B, Haberle J, Koch HG, Gortner L, Kreuder JG. Misleading diagnosis of partial N-acetylglutamate synthase deficiency based on enzyme measurement corrected by mutation analysis. *Acta Paediatr*. 2005;94:121-4.
42. Haberle J, Denecke J, Schmidt E, Koch HG. Diagnosis of N-acetylglutamate synthase deficiency by use of cultured fibroblasts and avoidance of nonsense-mediated mRNA decay. *J Inherit Metab Dis*. 2003;26:601-5.
43. Caldovic L, Morizono H, Panglao MG, Cheng SF, Packman S, Tuchman M. Null mutations in the N-acetylglutamate synthase gene associated with acute neonatal disease and hyperammonemia. *Hum Genet*. 2003;112:364-8.
44. Takanashi J, Barkovich AJ, Cheng SF, et al. Brain MR imaging in neonatal hyperammonemic encephalopathy resulting from proximal urea cycle disorders. *Am J Neuroradiol*. 2003;24: 1184-7.
45. Elpeleg O, Shaag A, Ben-Shalom E, Schmid T, Bachmann C. N-acetylglutamate synthase deficiency and the treatment of hyperammonemic encephalopathy. *Ann Neurol*. 2002;52:845-9.
46. Forget PP, van Oosterhout M, Bakker JA, Wermuth B, Vles JS, Spaapen LJ. Partial N-acetyl-glutamate synthetase deficiency masquerading as a valproic acid-induced Reye-like syndrome. *Acta Paediatr*. 1999;88:1409-11.
47. Plecko B, Erwa W, Wermuth B. Partial N-acetylglutamate synthetase deficiency in a 13-year-old girl: diagnosis and response to treatment with N-carbamylglutamate. *Eur J Pediatr*. 1998;157:996-8.
48. Morris AA, Richmond SW, Oddie SJ, Pourfarzam M, Worthington V, Leonard JV. N-acetylglutamate synthetase deficiency: favourable experience with carbamylglutamate. *J Inherit Metab Dis*. 1998;21:867-8.
49. Hinnie J, Colombo JP, Wermuth B, Dryburgh FJ. N-Acetylglutamate synthetase deficiency responding to carbamylglutamate. *J Inherit Metab Dis*. 1997;20:839-40.
50. Vockley J, Vockley CM, Lin SP, et al. Normal N-acetylglutamate concentration measured in liver from a new patient with N-acetylglutamate synthetase deficiency: physiologic and biochemical implications. *Biochem Med Metab Biol*. 1992;47:38-46.
51. Burlina AB, Bachmann C, Wermuth B, et al. Partial N-acetylglutamate synthetase deficiency: a new case with uncontrollable movement disorders. *J Inherit Metab Dis*. 1992; 15:395-8.
52. Pandya AL, Koch R, Hommes FA, Williams JC. N-acetylglutamate synthetase deficiency: clinical and laboratory observations. *J Inherit Metab Dis*. 1991;14:685-90.
53. Elpeleg ON, Colombo JP, Amir N, Bachmann C, Hurvitz H. Late-onset form of partial N-acetylglutamate synthetase deficiency. *Eur J Pediatr*. 1990;149:634-6.
54. Bachmann C, Brandis M, Weissenbarth-Riedel E, Burghard R, Colombo JP. N-acetylglutamate synthetase deficiency, a second patient. *J Inherit Metab Dis*. 1988;11:191-3.
55. Schubiger G, Bachmann C, Barben P, Colombo JP, Tonz O, Schupbach D. N-acetylglutamate synthetase deficiency: diagnosis, management and follow-up of a rare disorder of ammonia detoxication. *Eur J Pediatr*. 1991;150:353-6.